## PLNT4610 BIOINFORMATICS

## FINAL EXAMINATION

13:30 p.m. to 15:30 p.m. Friday December 9, 2011

Answer any combination of questions totalling to <u>exactly</u> 100 points. The questions on the exam sheet total to 120 points. If you answer questions totalling more than 100 points, answers will be discarded at random until the total points equal 100. This exam is worth 20% of the course grade.

Hand in the question sheets along with your exam booklet. All questions must be answered in the exam book. The question sheets will be shredded after the exam.

1. (5 points) What is an outgroup and why is it important to include one or more outgroups in phylogenetic analysis?

2. (10 points) Explain why the method of maximum parsimony is sensitive to the order of sequences in the dataset. What approach is usually taken to avoid this problem?

3. (10 points) In eukaryotes, why are the polymorphisms seen with molecular markers usually selectively neutral?

4. (10 points) Compare and contrast the use of molecular markers, DNA sequences, and protein sequences for constructing phylogenies:

- for different populations of the same species
- for different species that are closely-related
- for different species that are distantly-related

5. (5 points) Genetic linkage maps were inferred, based on segregation of molecular markers. The names of three maps and their log likelihoods are listed below. Which map is the map most likely to have given rise to the observed data:

	log likelihood
Map A	- 124.30
Map B	- 205.01
Map C	-377.66

6. (10 points) In many microarray systems, it is now common practice to "spike" each RNA sample with a set of well-quantified synthetic RNAs, which get labeled as cDNA along with the rest of the RNA population. Oligonucleotides complementary to the spike RNAs are also included on the array. Explain the function of the RNA spikes.



7. (15 points) Design of microarray experiments makes the distinction between biological replicates and technical replicates. In biological replicates, the entire biological experiment is repeated, and new RNA samples extracted from each experiment, and each sample is labeled and hybridized independently to different microarrays. In technical replicates, the same RNA from a given experiment is labeled in separate labeling reactions, and hybridized to different microarrays. Explain what biological replicates and technical replicates tell you. Which is more important and why?

8. (10 points) Two methods for calculating pairwise distances between DNA sequences are the Jukes and Cantor method and the Kimura 2-parameter method. In Jukes & Cantor, the rate of base substitution is assumed to be the same for all possible base substitutions (eg. A to G, A to C, T to G etc.) The Kimura 2-parameter method weights transversions (purine to pyrimidine or pyrimidine to purine) more strongly than transitions (ie. purine to purine or pyrimidine to pyrimidine substitutions). This is in agreement with the observation that transitions occur more frequently than transversions. Usually, transversions are weighted twice as much as transitions.

If distance matrices are constructed for the same sequence alignment, using either the Jukes and Cantor method, or the Kimura 2-parameter method, what differences would you expect for Neighbor Joining trees constructed using the two different matrices?

9. (10 points) The spreadsheet below shows data for a set of molecular markers for 12 individuals in a population. The rows list the names of the 12 individuals. The columns B - U represent presence or absence of a band for each of 20 markers scored for each individual.

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1	A001	0	1	1	1	1	1	0	1	1	0	1	1	0	1	1	1	1	1	0	1	
2	A002	0	0	0	1	0	0	0	0	1	1	0	0	0	0	1	0	0	1	0	0	
3	A003	0	0	1	1	1	1	0	1	1	0	1	1	0	1	1	1	1	1	0	1	Ξ
4	A004	0	1	1	1	1	0	0	1	1	0	0	1	1	1	1	1	1	1	0	1	
5	A005	0	1	1	1	1	1	0	1	1	0	1	1	0	0	0	1	0	1	0	1	
6	A006	0	0	1	1	1	1	0	1	1	0	0	1	0	0	1	1	1	1	0	1	
7	A007	0	1	1	1	1	0	0	0	1	0	0	1	0	0	1	1	1	1	0	1	
8	A008	0	1	1	1	1	0	1	0	1	0	1	1	0	1	1	1	1	1	0	0	
9	A009	0	1	1	1	1	0	0	1	1	0	1	1	0	1	1	1	1	1	0	0	
10	A010	0	1	1	1	1	0	0	1	1	0	0	1	0	0	1	1	1	1	1	0	
11	A011	0	1	1	1	1	0	0	1	1	0	1	1	1	1	1	1	1	1	0	0	
12 A012 0 1 1 1 0 0 1 1 0 1 1 0 1 1 1 0 0 •																						
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Are some markers in this dataset more informative than others? Explain.

10. (10 points) Ontologies are a formalized method for describing concepts. Ontologies are therefore an essential first step in the long term goal of computer "reasoning". For any biological concept you wish, draw an ontology diagram. It does not need to be elaborate, but it should describe the relations between different levels of objects.

11. (10 points) In phylogenetic analysis of molecular marker data, programs that perform distance and maximum likelihood methods require the user to specify a "Site length". What does this term refer to, and why is it important?

12. (15 points) Based on the GenBank flat file entry on the next page, draw a schema for the database. The objective is to use a small number of well designed classes that cleanly describe the components of the data and their relationships. (Note: For brevity, only the first few lines of the sequence are shown below.)

LOCUS	NM_001082679 1619 bp mRNA linear MAM 05-DEC-2010
DEFINITION	Oryctolagus cuniculus coagulation factor VII (serum prothrombin
	conversion accelerator) (F7), mRNA.
ACCESSION	NM_001082679
VERSION	NM_001082679.1 GI:130495947
KEYWORDS	
SOURCE	Oryctolagus cuniculus (rabbit)
ORGANISM	Oryctolagus cuniculus
	Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi;
	Mammalia; Eutheria; Euarchontoglires; Glires; Lagomorpha;
	Leporidae; Oryctolagus.
REFERENCE	1 (bases 1 to 1619)
AUTHORS	Wong,P.C., Luettgen,J.M., Rendina,A.R., Kettner,C.A., Xin,B.,
	Knabb,R.M., Wexler,R. and Priestley,E.S.
TITLE	BMS-593214, an active site-directed factor VIIa inhibitor: enzyme
	kinetics, antithrombotic and antihaemostatic studies
JOURNAL	Thromb. Haemost. 104 (2), 261-269 (2010)
PUBMED	20589312
REMARK	GeneRIF: Results suggest that inhibition of FVIIa with
	small-molecule active-site inhibitors represents a promising
	antithrombotic approach.
REFERENCE	2 (bases 1 to 1619)
AUTHORS	Brothers, A.B., Clarke, B.J., Sheffield, W.P. and Blaichman, M.A.
TITLE	Complete nucleotide sequence of the cDNA encoding rabbit
	coordination factor VII
JOURNAL	Thromb. Res. 69 (2), 231-238 (1993)
PUBMED	8383365
COMMENT	PROVISIONAL REESED: This record has not yet been subject to final
CONTRACTOR	NCBI review. The reference sequence was derived from U77477.1.
FFATURES	Location/Qualifiers
sourc	e 11619
00010	/organism="Oryctolagus_cuniculus"
	/organism of jorgingus confidents
	/db_xref="taxon:9986"
nene	1 1619
gene	/gene="E7"
	/pote="coagulation_factor_VII_(serum_prothrombin
	conversion accelerator)"
	/db xref="GeneID:100009399"
CDS	221356
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	KTDHDTALLEN OPAALTNNV/PLOLPERNESESTLATTRESRVSGWGOLLYRGALAP
	GTWYLTGVVSWGEGCAAVGHVGVYTRVSRYTEWLSRLMRSKLHHGTORHPEP"
	STWIETOVOWSEGOARONVONVONVOVITEWESKERKOKENIOTQKII II
L 61	ctonenetee aanontetet nnetheante tttataacee annannae neacanente
101	ctararanar aaaaaraaaar raattette etaaaaaan taraaraaaa etaastaaaa
121	οιγογοαγγο απαγγογγγο σααιτοιιτο σιγγαγγάγο ιγογγουγγγ στογοιγγάγ

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