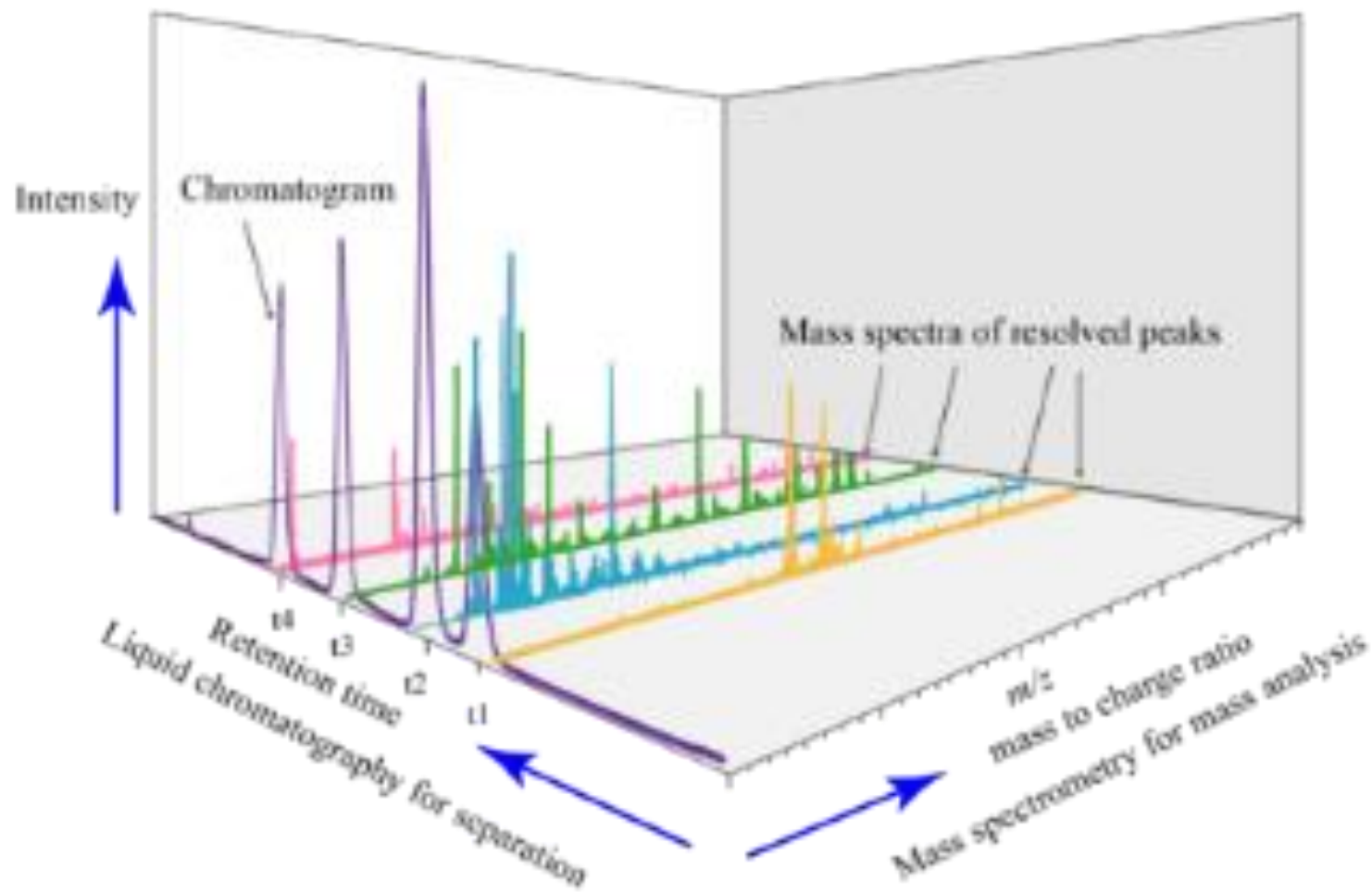


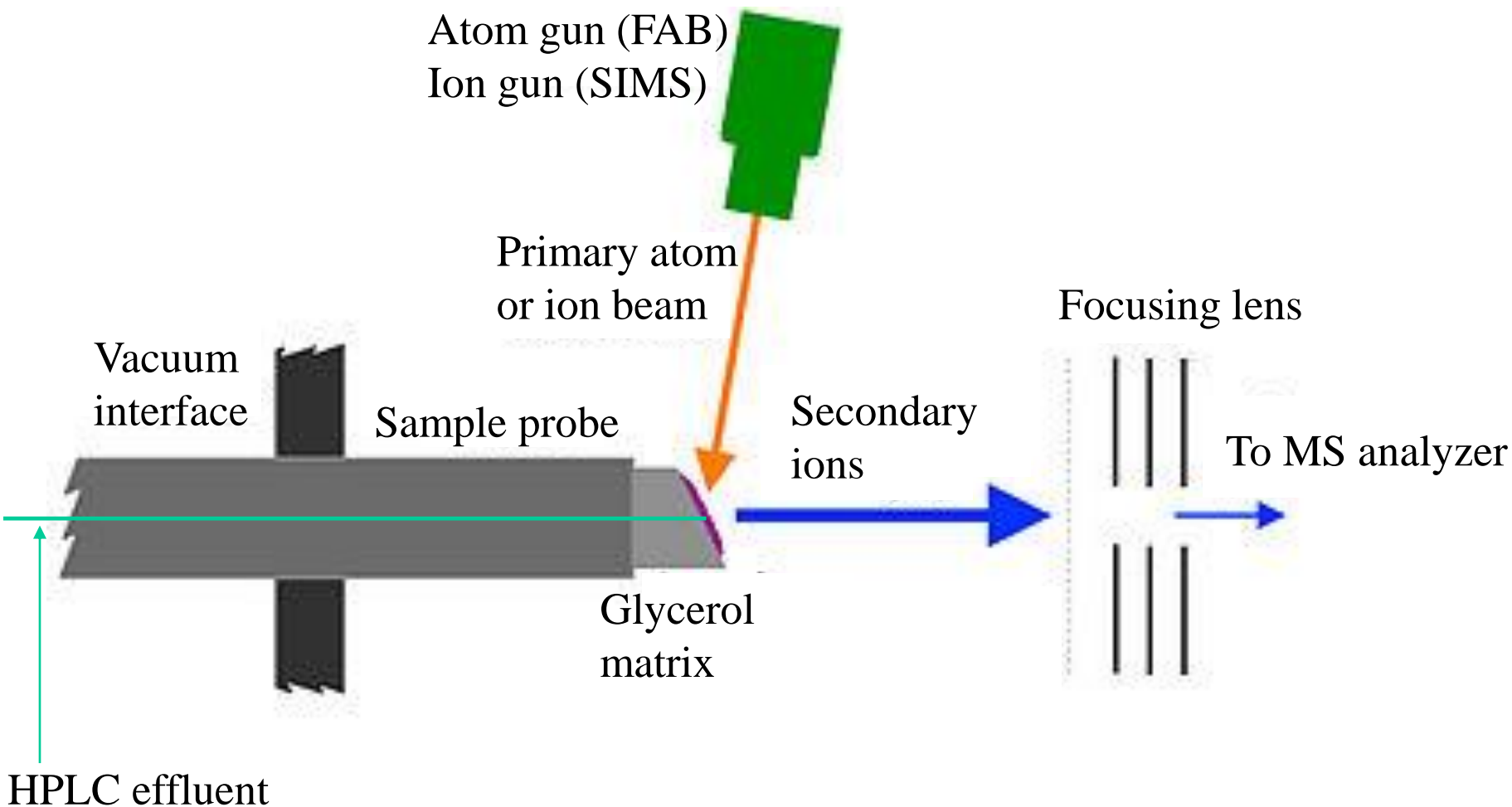
HPLC-MS

LC-MS interfaces

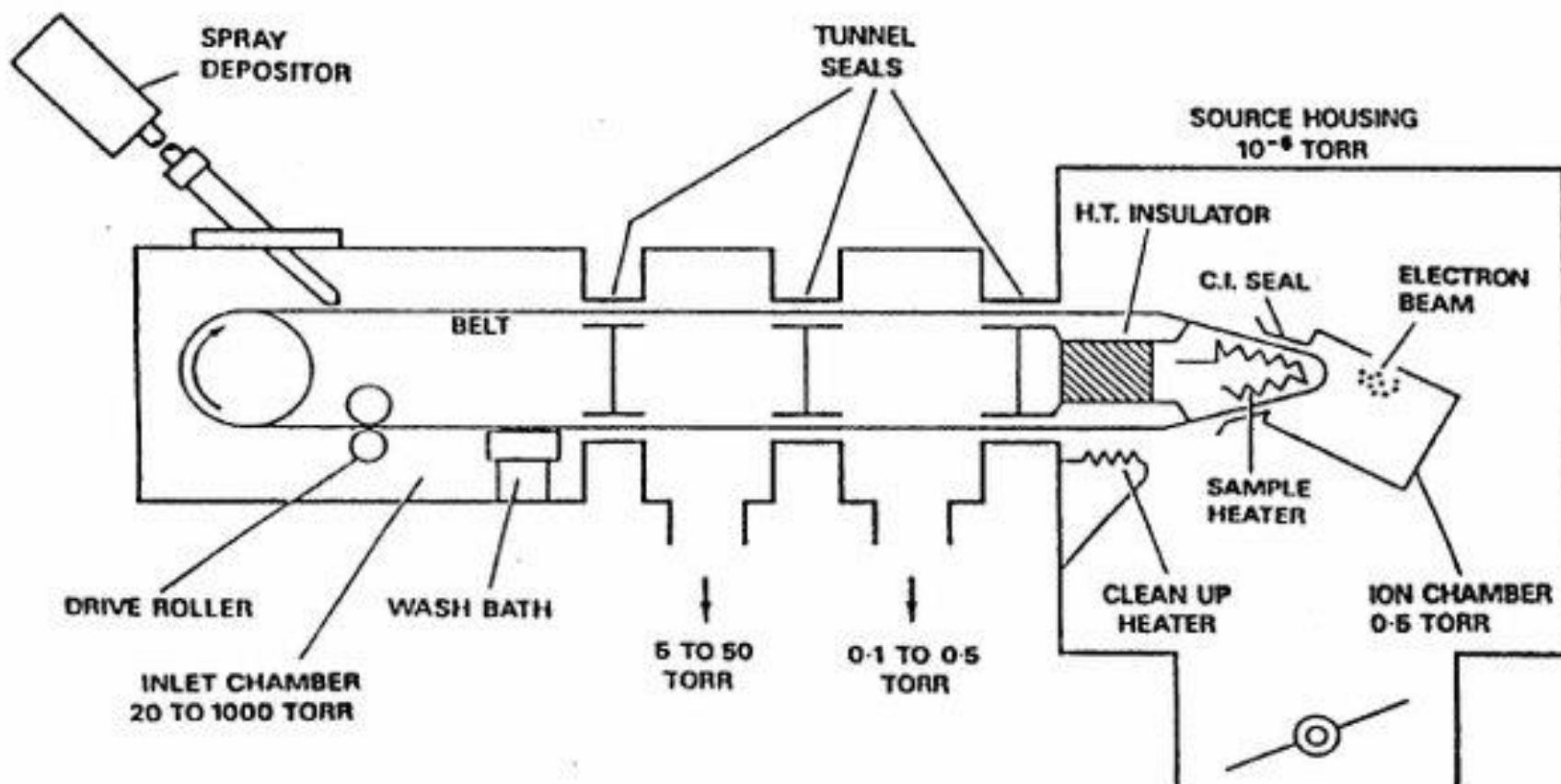
- Continuous-flow secondary ion mass spectrometry (SIMS)
Continuous-flow fast atom bombardment (FAB)
- Moving belt interface
- Atmospheric pressure chemical ionization (APCI)
- Electrospray ionization (ESI)



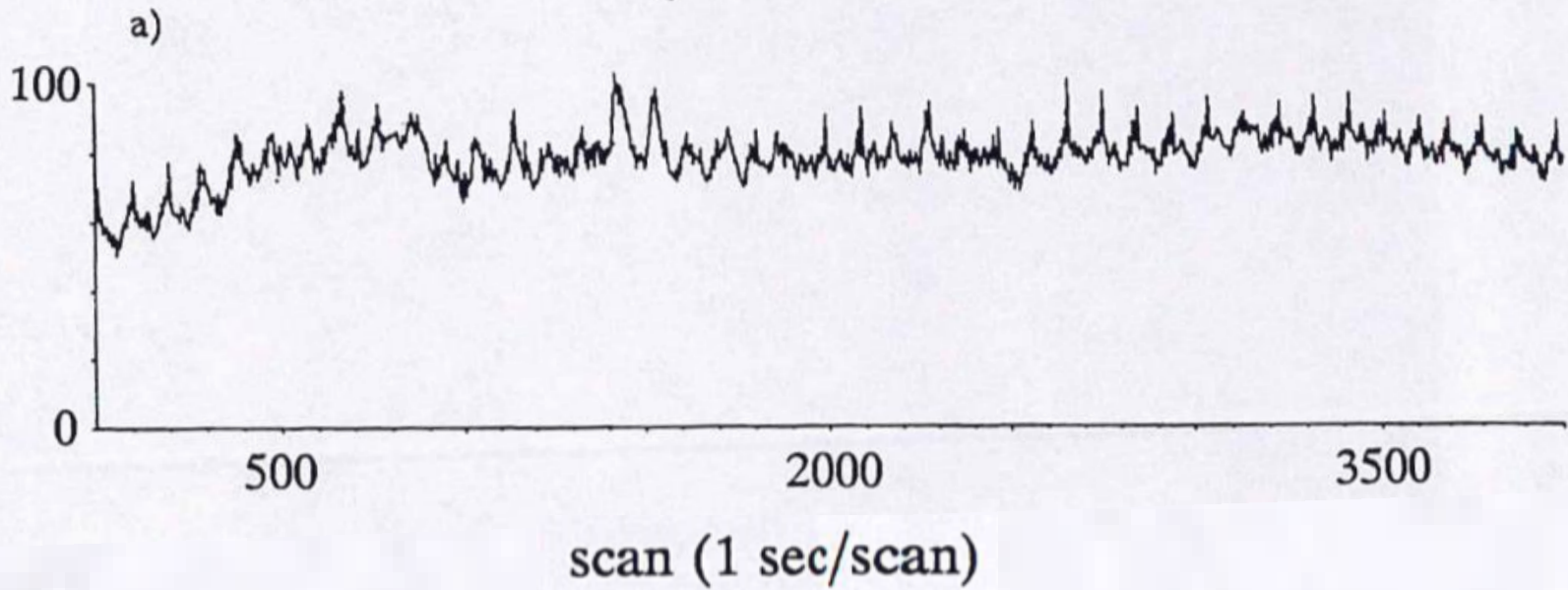
Continuous-flow SIMS or FAB

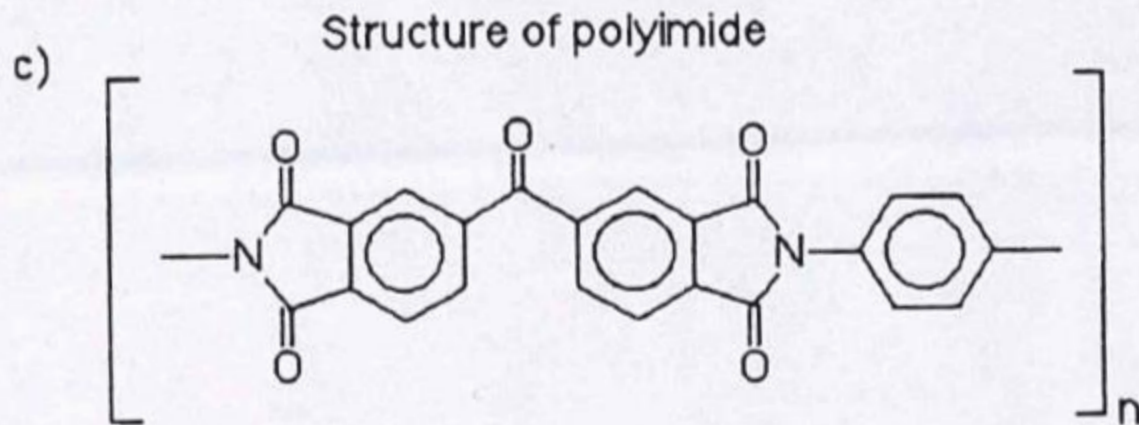
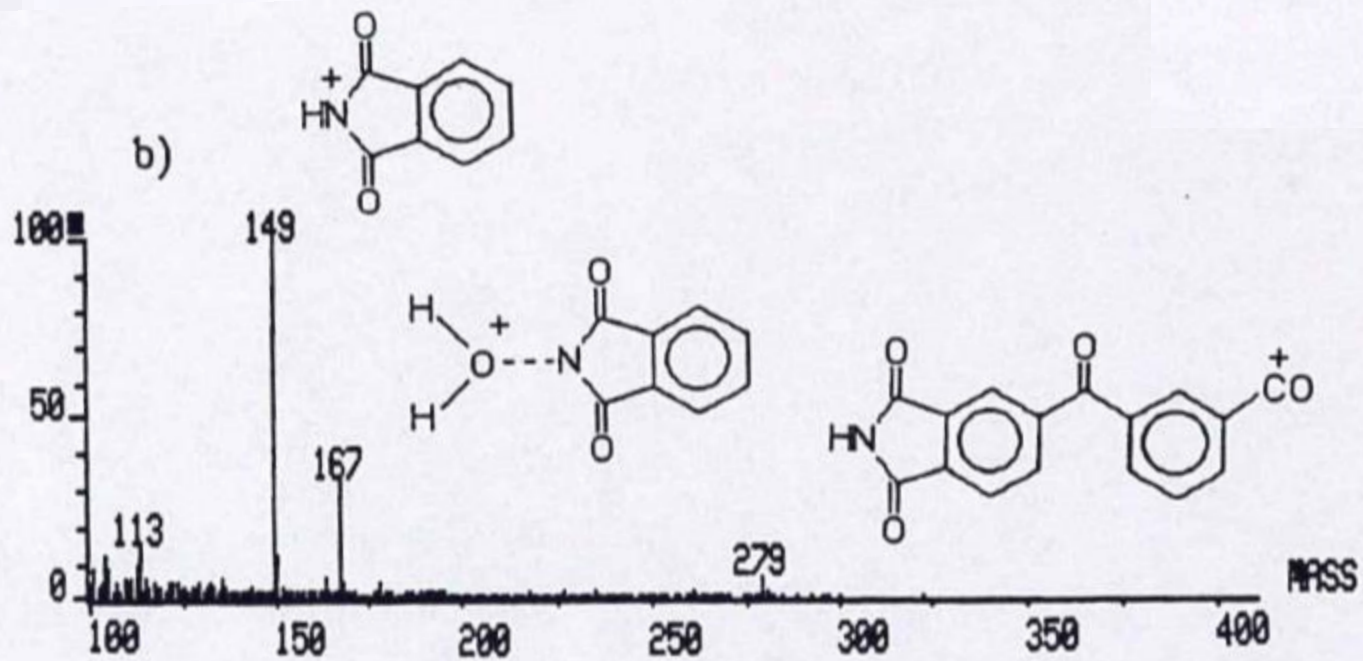


Moving belt interface for EI/CI-MS

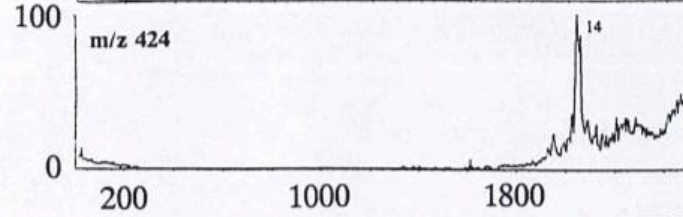
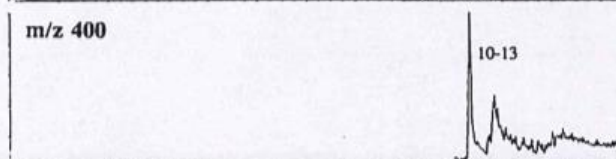
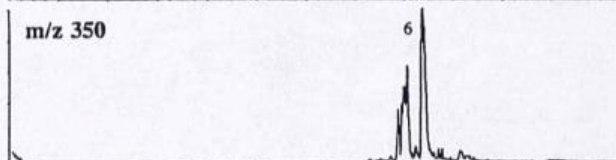
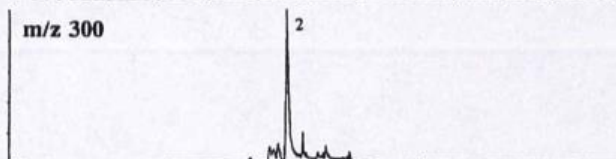
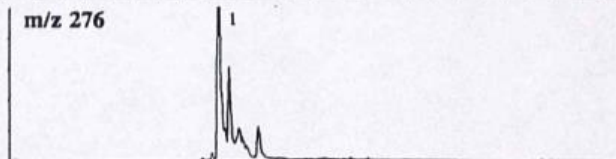
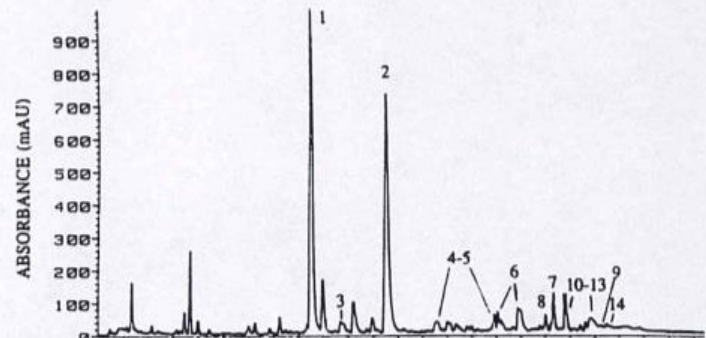
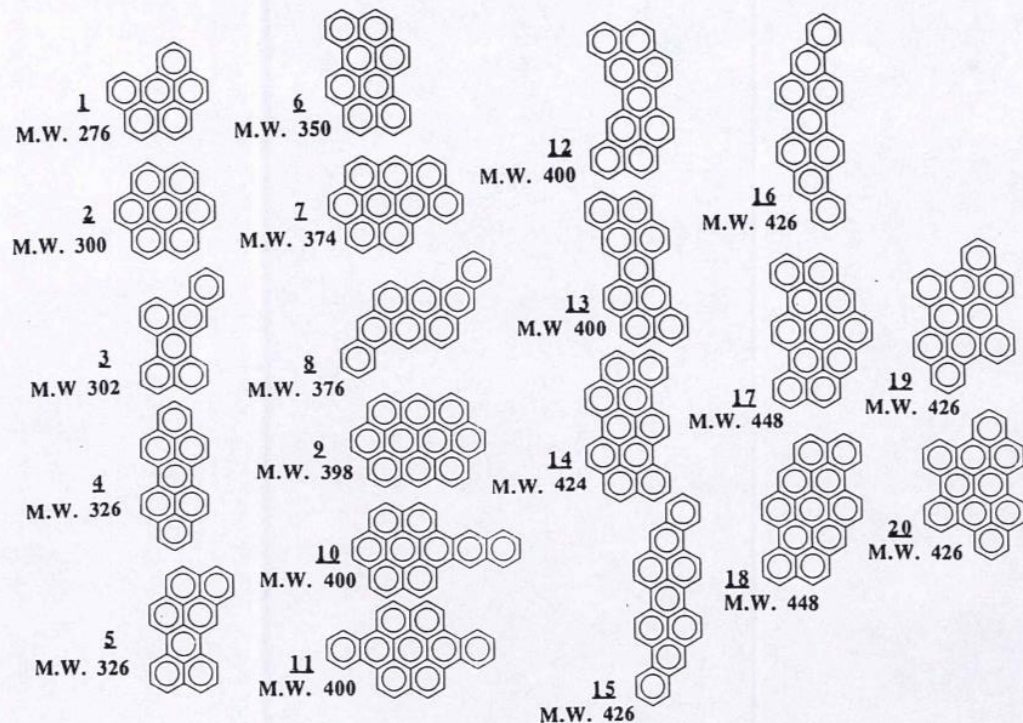


Total ion current, belt only



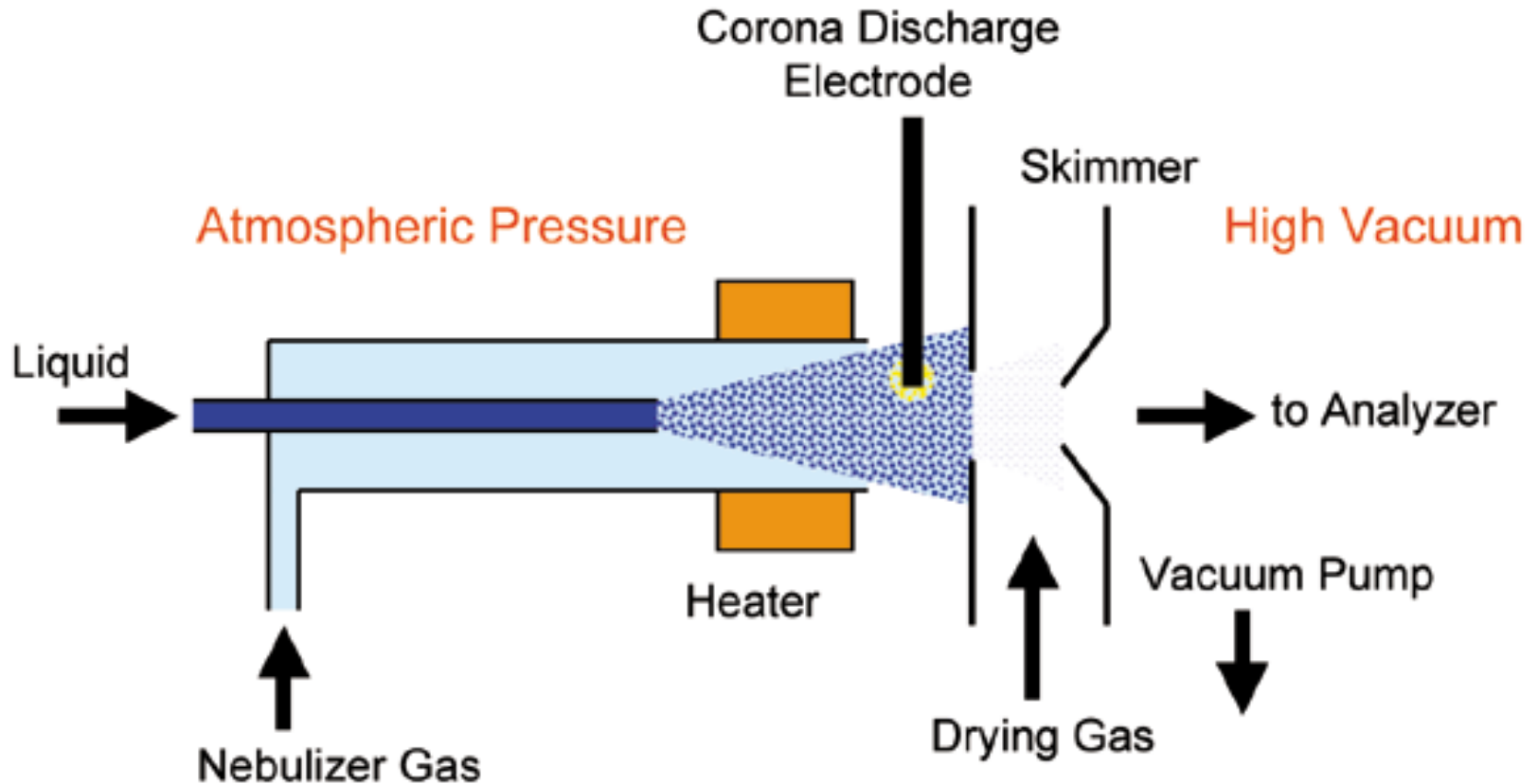


UV (254 nm) and SIC for PACs

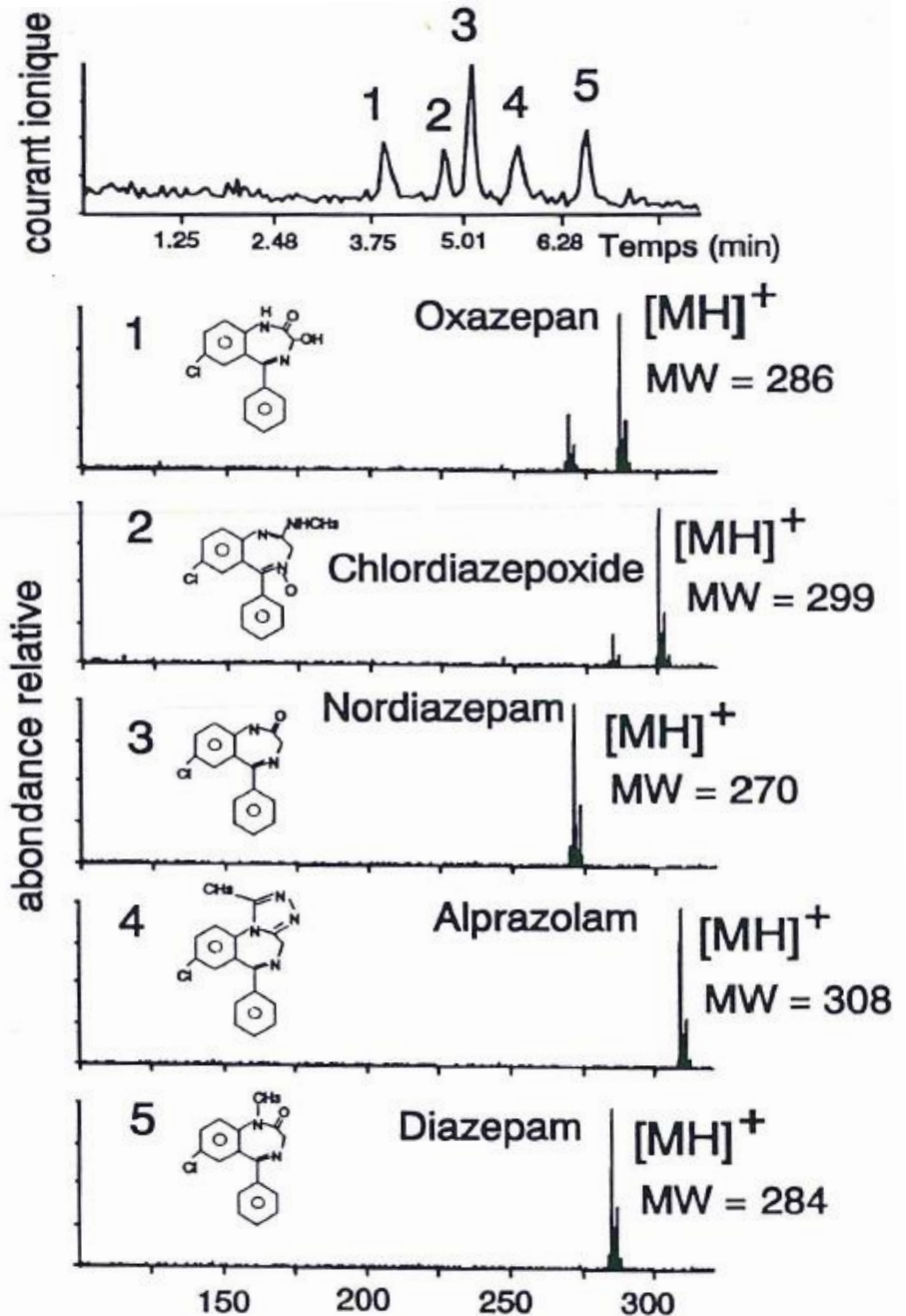


scan (3 sec/scan)

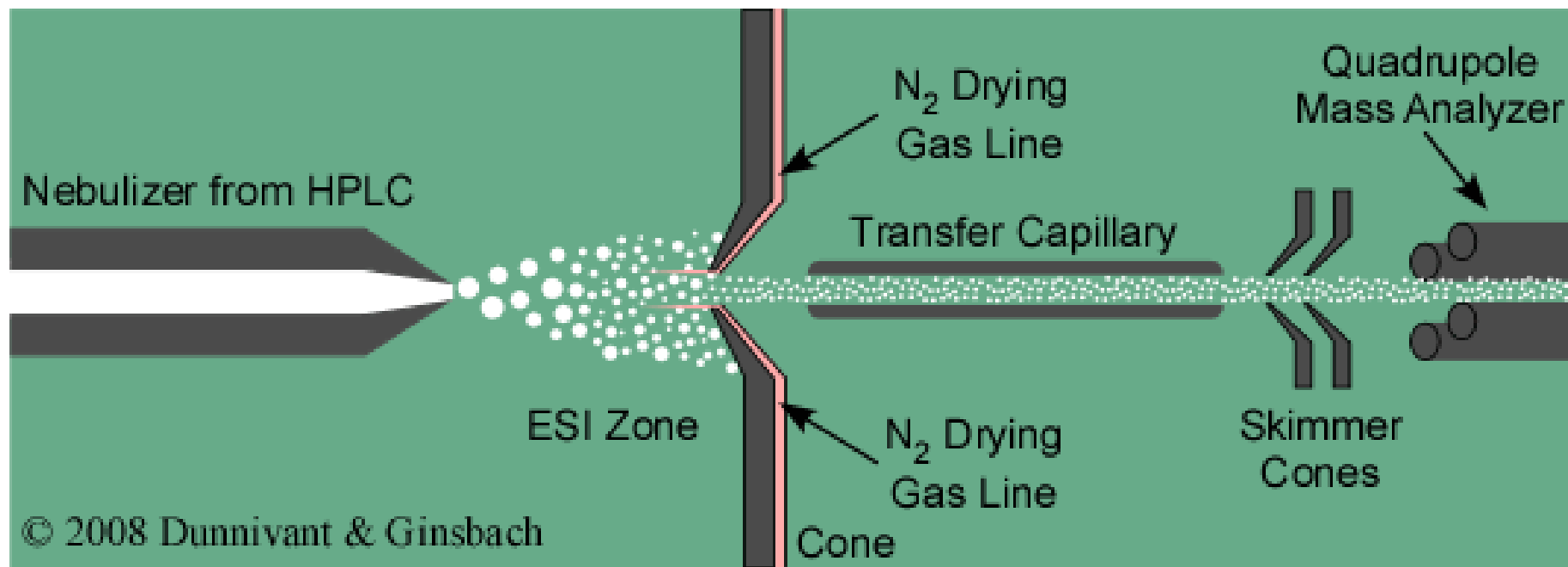
Atmospheric pressure CI interface



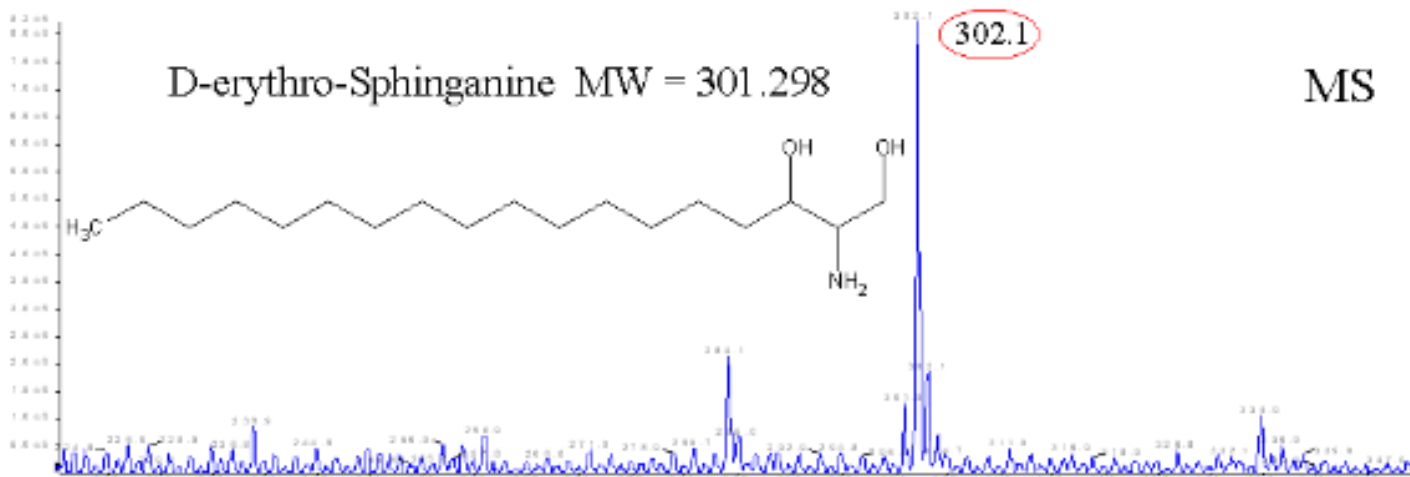
RP-HPLC/MS of benzodiazepines (APCI)



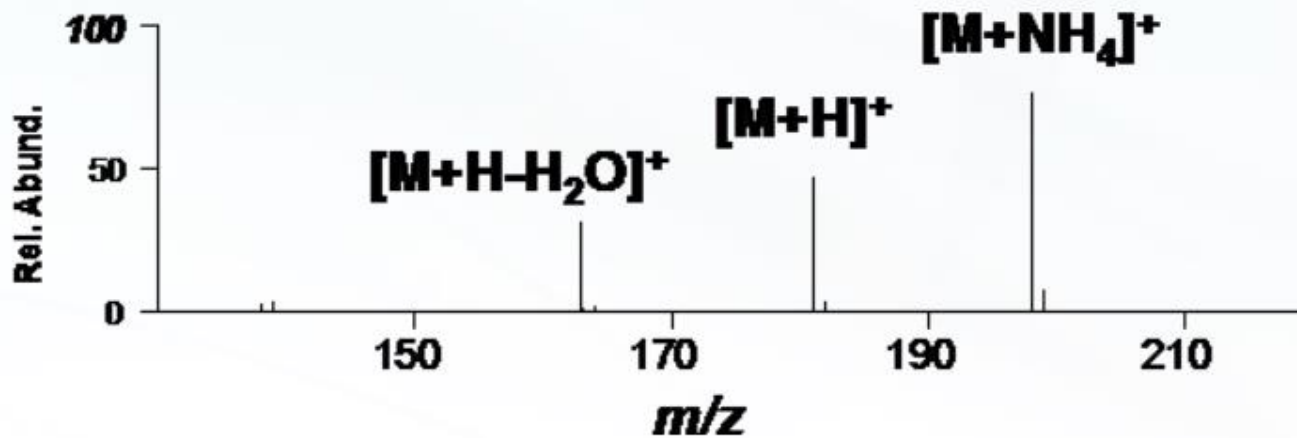
Electrospray ionization interface



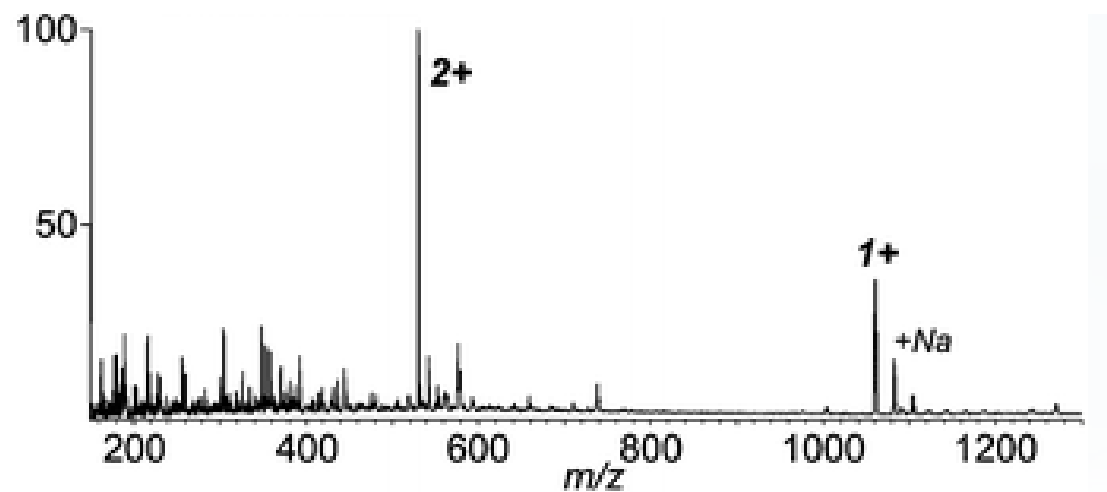
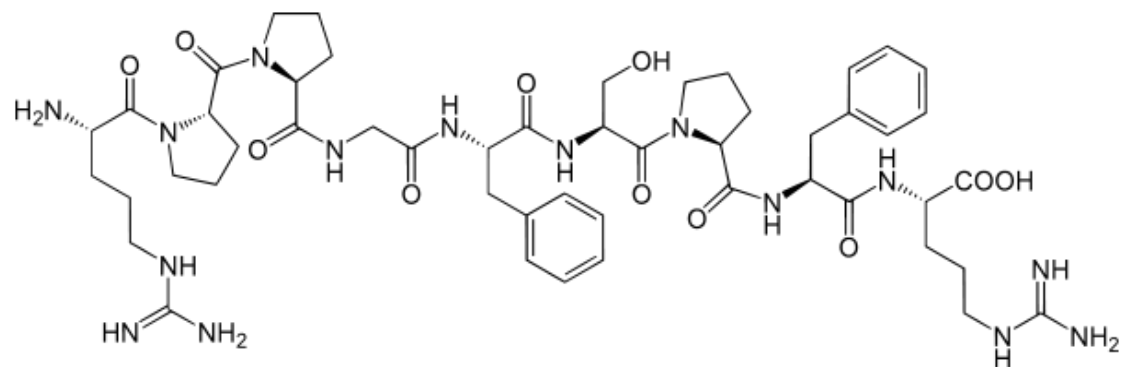
ESI-MS of small molecules



Aspirin



ESI-MS of larger molecules



ESI-MS of a large protein

hen egg lysozyme

Platform II, BMB, The University of Leeds

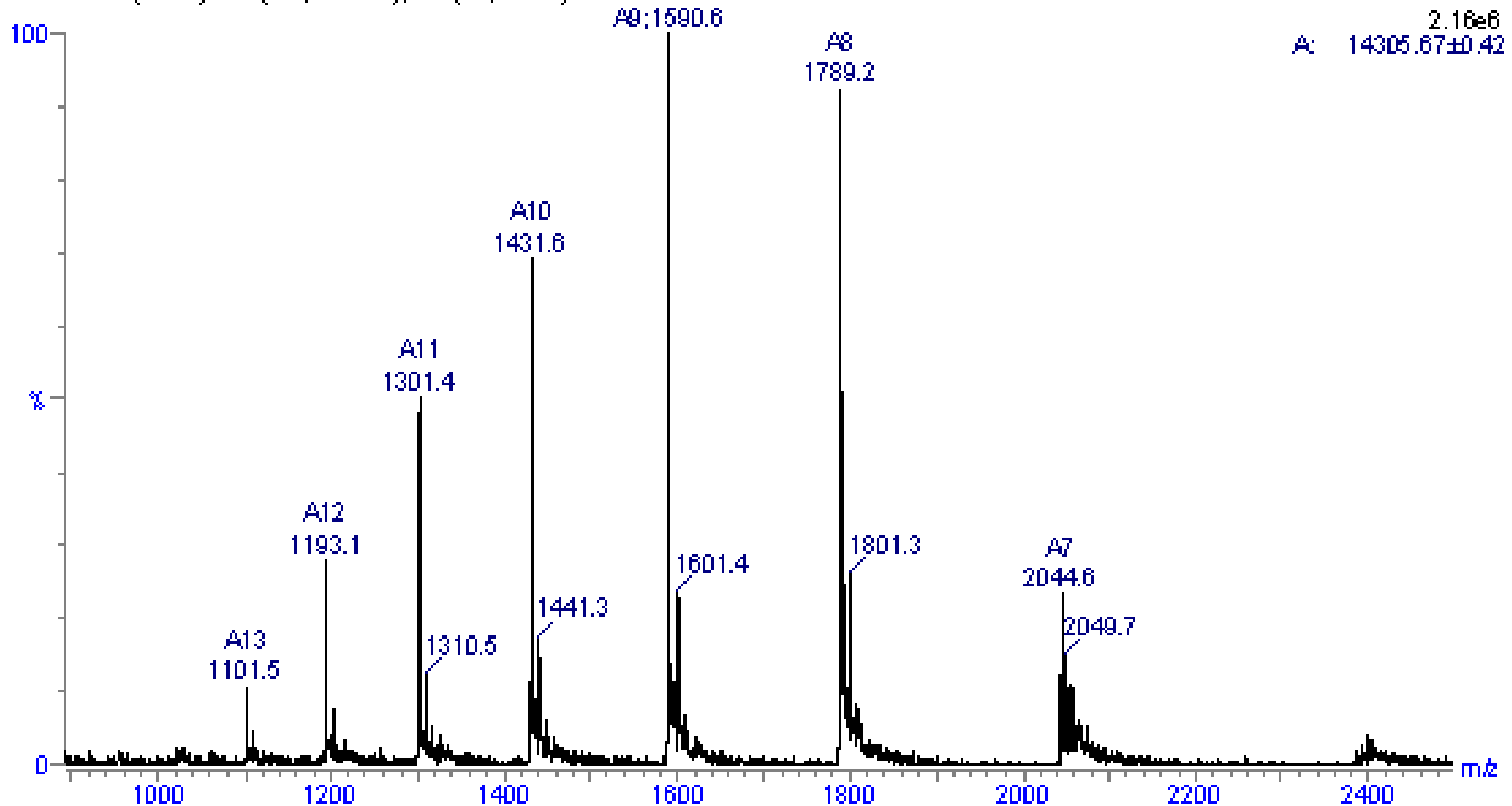
25-Jan-2000 10:00:37

LYSD1A_1 (1.392) Sm (SG, 2x1.00); Sb (10,10.00)

Scan ES+

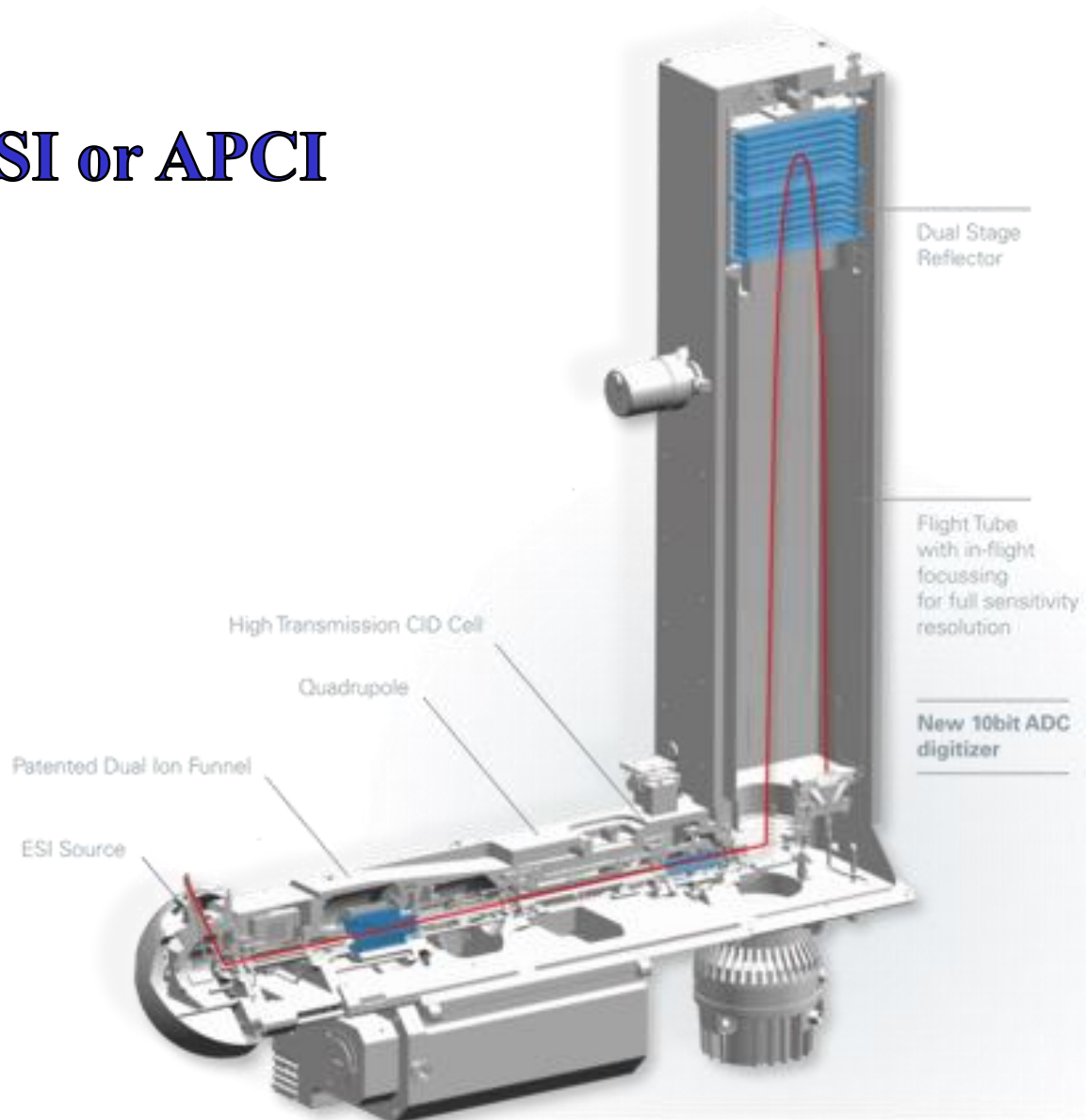
2.16e6

A: 14305.67±0.42



In MCAL: quadrupole-time-of-flight tandem analyzer

ESI or APCI



Anal Bioanal Chem (2010) 398:769–777

DOI 10.1007/s00216-010-3931-1

ORIGINAL PAPER

Application of LC–TOF MS to analysis of hemoglobin acetaldehyde adducts in alcohol detoxification patients

**Stefan W. Toennes • Moritz G. Wagner •
Gerold F. Kauert**

RATIONALE:

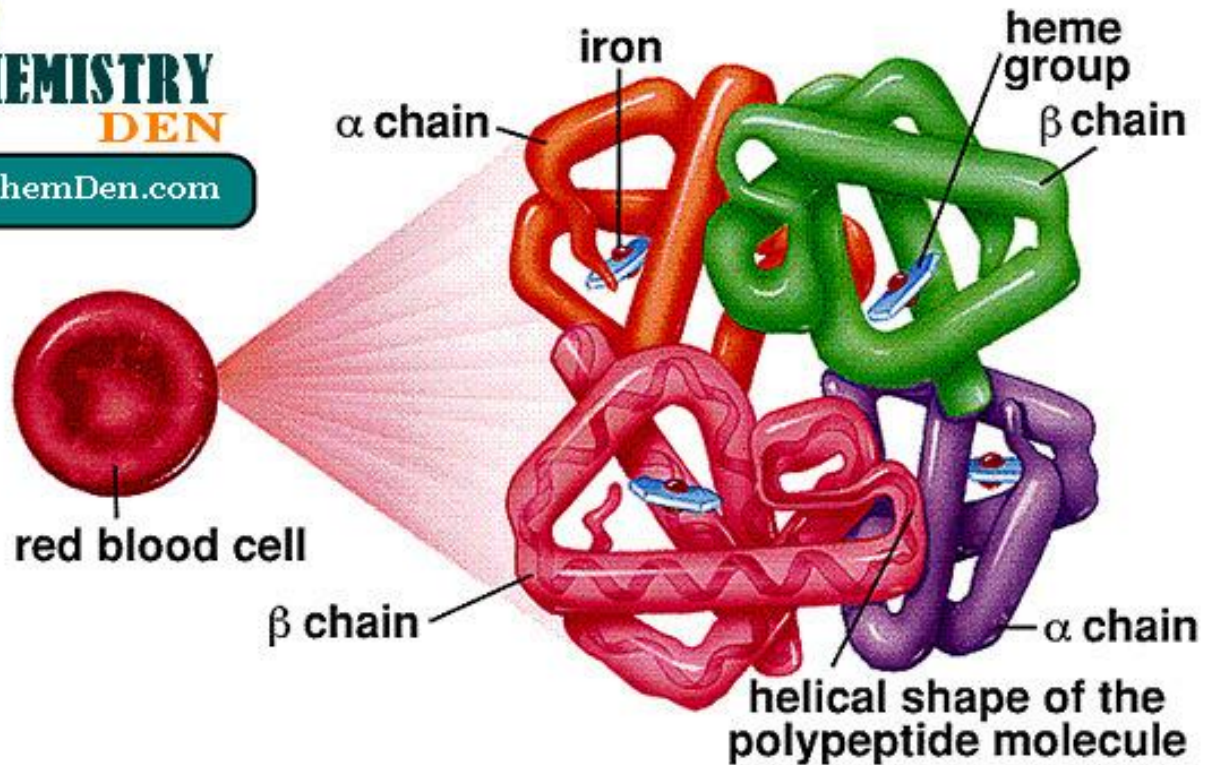
- Acetaldehyde, as first oxidative metabolite of ethanol, binds *in vivo* to a wide variety of molecules, e.g. to albumin, low density lipoproteins, ribonuclease, nucleosides, and hemoglobin (Hb).
- Acetaldehyde adducts of hemoglobin are regarded as potential markers of ethanol consumption.
- **Hemoglobin** from blood samples of alcohol addicts before and during detoxification was obtained, analyzed and compared with hemoglobin taken from children.

HEMOGLOBIN MOLECULE



BIOCHEMISTRY
DEN

www.BiochemDen.com



<https://www.biochemden.com/hemoglobin/>

EXPERIMENTAL:

Red blood cells were lysed and filtered.

Hemolysate (200 μ L, containing approximately 20 mg of Hb) was mixed with an equal volume of 80:20 acetonitrile–water containing 0.2% TFA to denature Hb.

This was centrifuged at 16000 g for 10 min.

Of the supernatant, 0.5 μ L was analyzed by LC–TOF MS using a linear gradient with 0.1% TFA in water as solvent A and acetonitrile as solvent B.

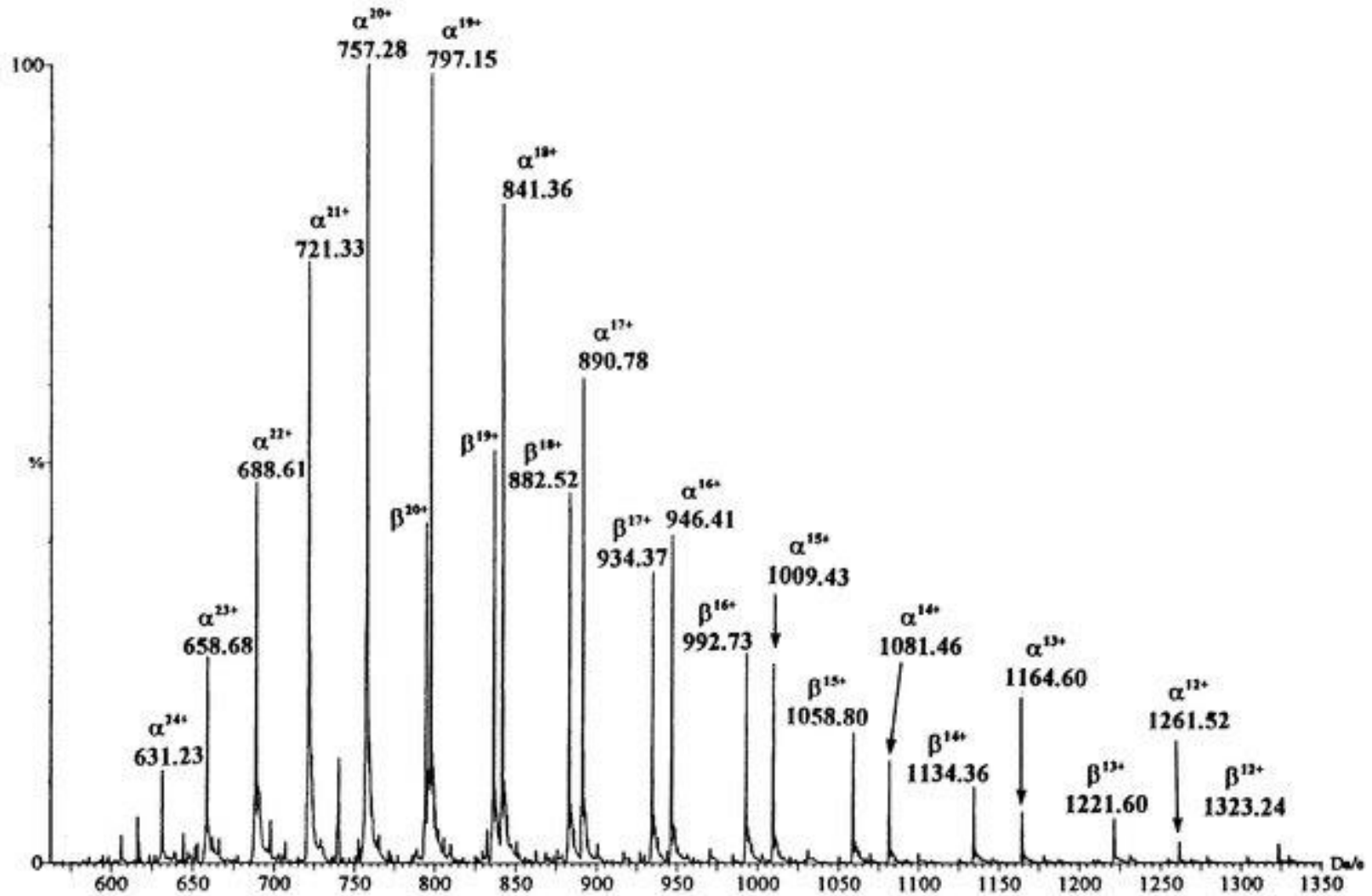
The % B was increased from 40% to 50% in 10 min, followed by a washing step with 100% B for 2 min.

The flow rate was 0.4 mL/min and the column was operated at 50 °C.

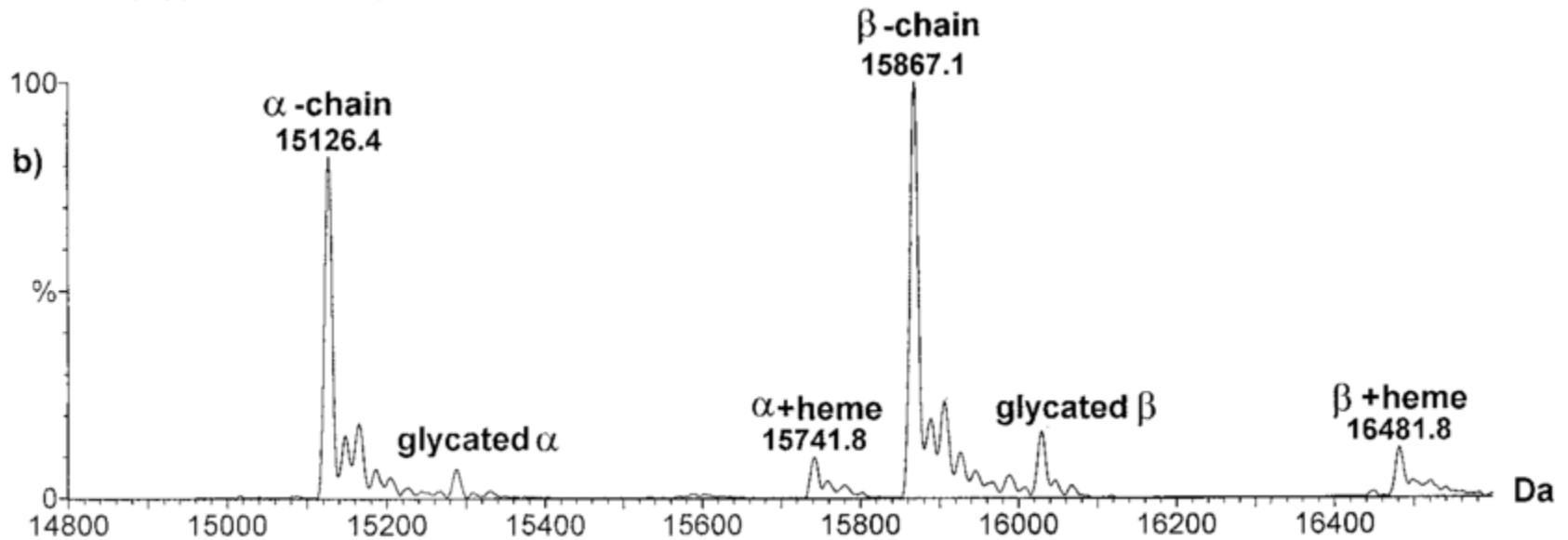
Column: C18-Ether column (100 \times 2.0 mm, 3 μ m particle size)

MS: ESI-TOF (Agilent)

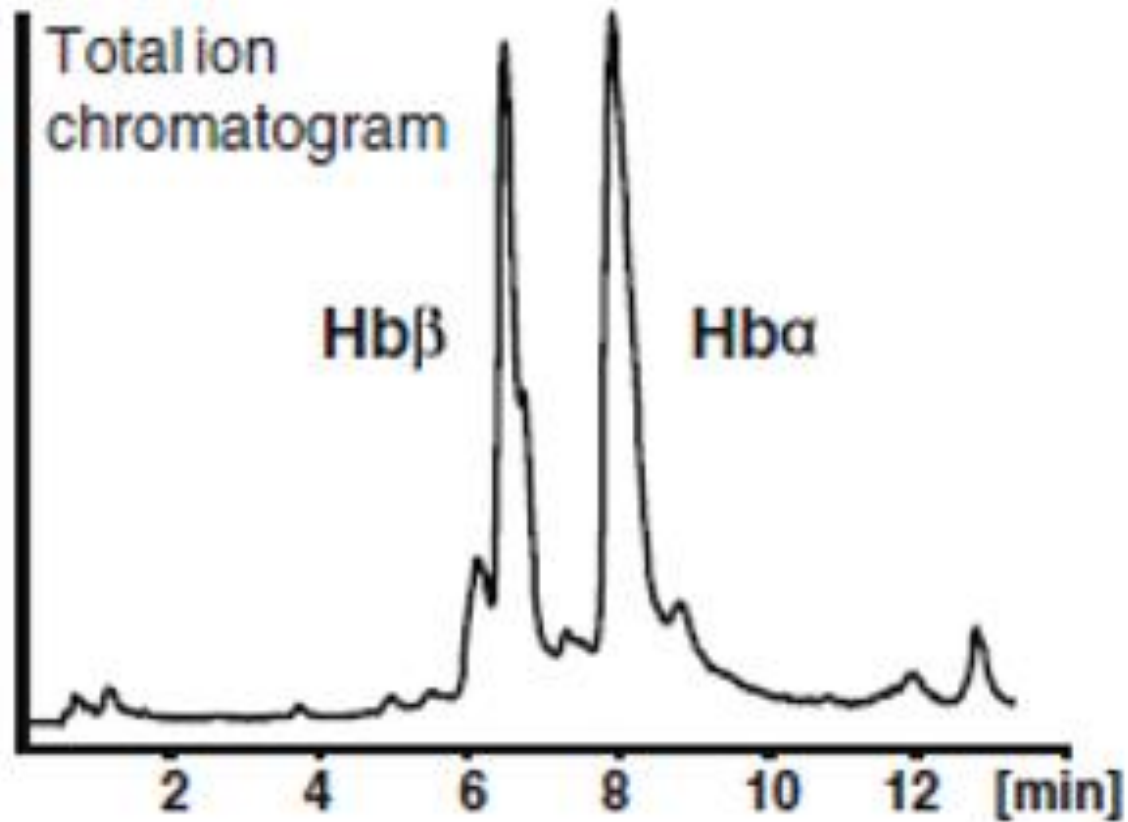
ESI-MS spectrum of normal Hb alpha + beta chains



Deconvoluted ESI-MS spectrum of normal Hb alpha + beta chains

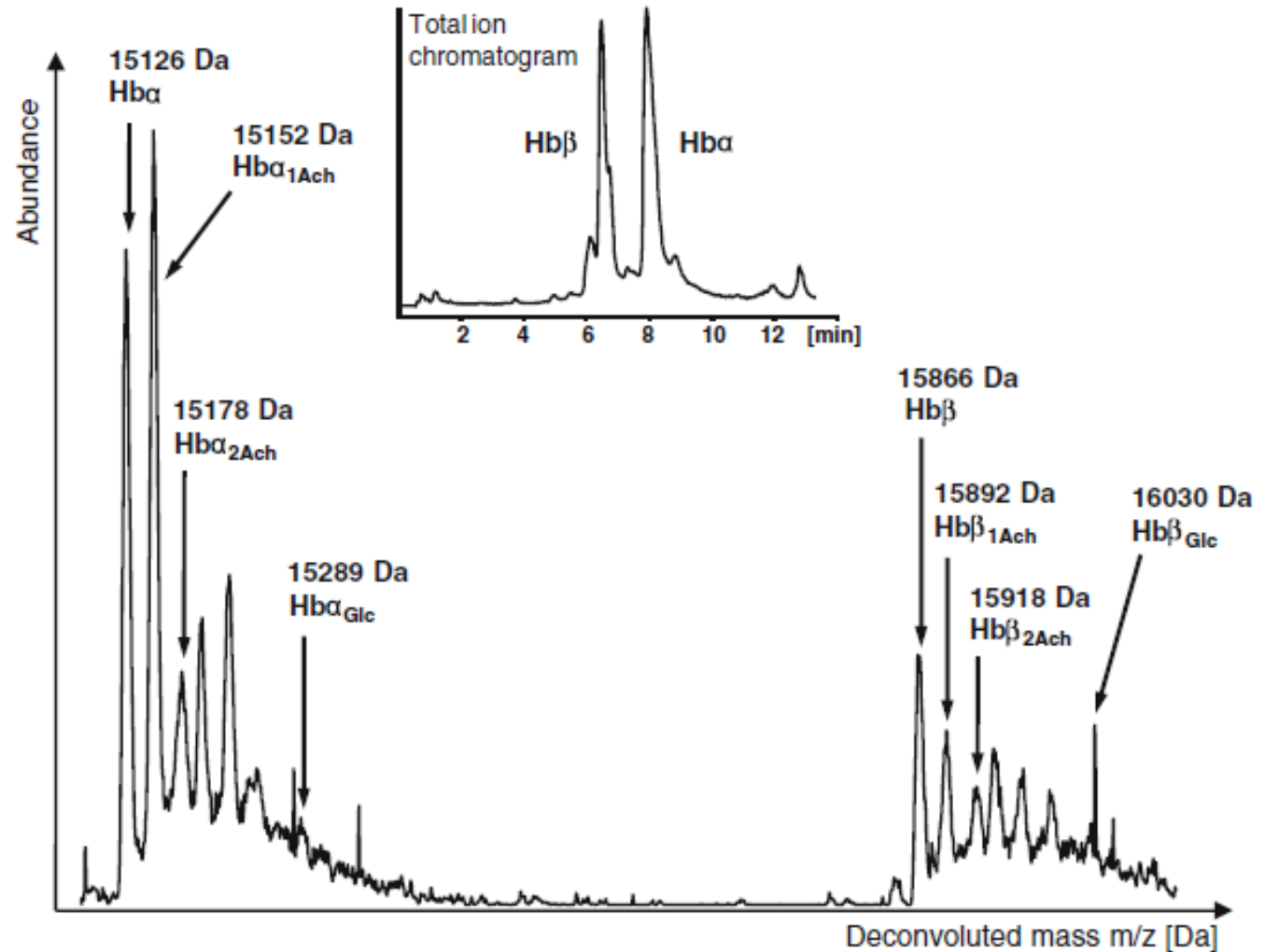


LC-MS TIC



Deconvoluted ESI mass spectrum averaged from 6-9 min range

Fig. 1 Deconvoluted mass peaks resulting from the mass spectra in the retention time range 6–9 min; the total-ion chromatogram of the hemoglobin chain separation is shown in the *inset*. The hemoglobin alpha ($Hb\alpha$) and beta ($Hb\beta$) chains and their modified homologues (glycosylation, *Glc*; and acetaldehyde modification, *Ach*) are annotated



**Agilent 6200 Series Accurate-Mass
Time-of-Flight (TOF) LC/MS**

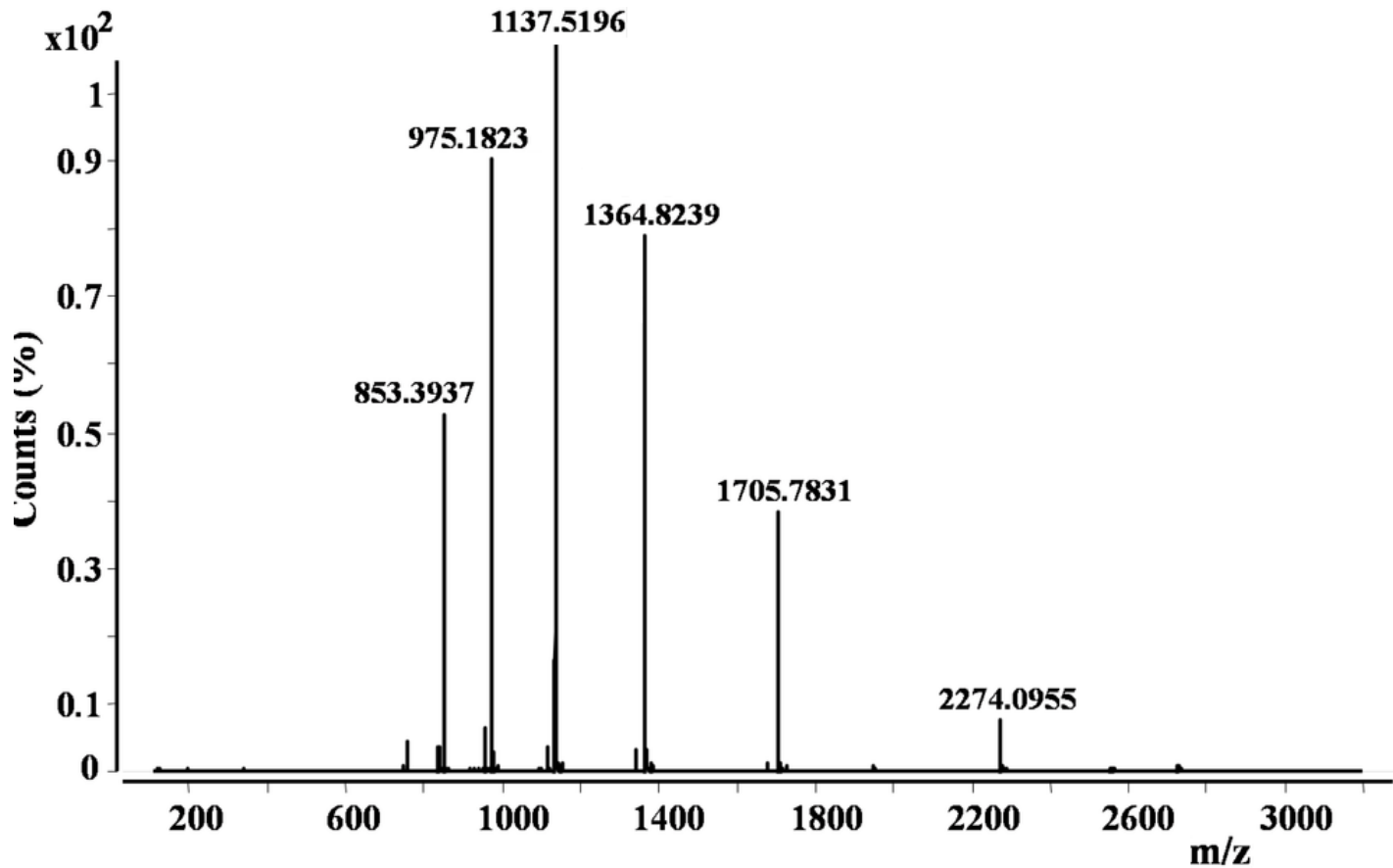


Summary LC-MS

- Good for liquid and solid samples
- All size of molecules
- Normal, reversed phase LC possible
- UV/fluorescence detector in series or parallel
- Very specific owing to mass measurement

Questions

1. Calculate the molecular mass of the compound that produced this positive mode electrospray spectrum:



2. a) In LC-MS, reversed-phase HPLC is much more used than normal phase HPLC. Discuss one possible reason for this fact.

b) In LC-MS why is the flow split after the column?