

PLNT2530 PLANT BIOTECHNOLOGY

MID-TERM EXAMINATION

11:30 am to 12:20 pm Monday, March 14, 2022

Answer any combination of questions totaling to exactly 100 points. If you answer questions totaling more than 100 points, answers will be discarded at random until the total points equal 100. The questions total to 120 points. This exam is worth 20% of the course grade.

Hand in these question sheets along with your exam book. Question sheets will be shredded.

Ways to write a readable and concise answer:

- i. Just answer the question. Save time by specifically addressing what is asked. Don't give irrelevant background if it doesn't contribute to the question that was asked.
 - ii. Avoid stream of consciousness. Plan your answer by organizing your key points, and then write a concise, coherent answer. Make your point once, clearly, rather than repeating the same thing several times with no new information.
 - iii. Point form, diagrams, tables, bar graphs, figures are welcome. Often they get the point across more clearly than a long paragraph.
 - iv. Your writing must be legible. If I can't read it, I can't give you any credit.
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1. (10 points) Why is it that plants cells in tissue culture can regenerate into complete organ systems such as shoots or roots? In particular, in what way do plants lend themselves to regeneration, where animals do not?

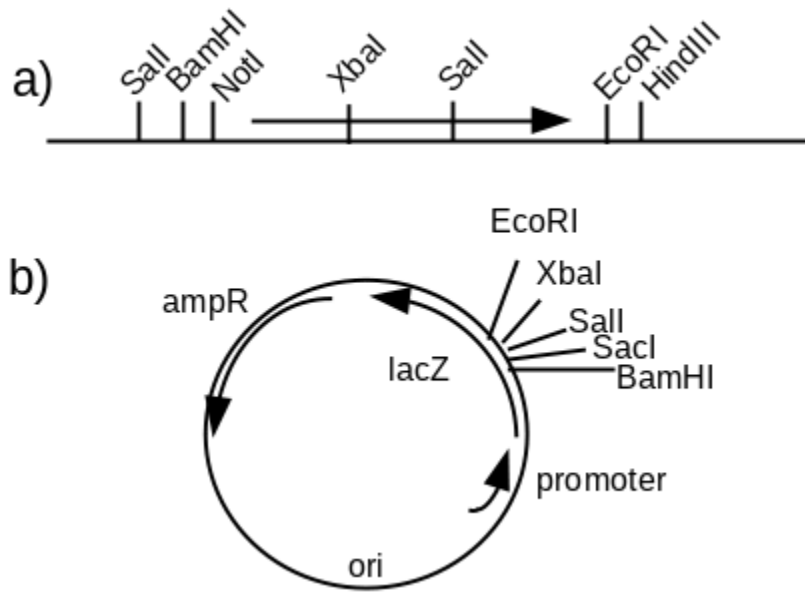
2. (10 points) When making a cDNA library with a Lambda phage vector, the protocol typically includes cutting with a restriction enzyme, followed by dephosphorylation of the vector using alkaline phosphatase. Alternatively, one might consider dephosphorylating the cDNA instead of the Lambda arms. What would be the problem with this alternative approach?

3. (5 points) The simplest way to construct an expression library would be to synthesize a double-stranded cDNA, make the ends blunt using DNA polymerase, and then ligate identical adaptors for a restriction enzyme to both ends. For example, if the adaptors had overhanging ends for a BamHI site, you could clone the cDNA into the BamHI site of a vector.

Suppose you screened the library using a hybridization probe, and found 24 positive clones. How many of them do you expect would express the protein?

Hint: This is NOT directional cloning.

4. (10 points) Based on the restriction maps for a gene (a) and a vector (b), choose a pair of restriction enzymes that could be used to cut the entire gene from a, to be cloned between any two compatible sites in the vector.



5. (10 points)

a) Imagine that you are testing a self-driving car on a street with 20 houses. In each trial, the computer chooses an address at random, and drives to that address. After 20 trials, would you expect to have visited all 20 houses? Explain your answer.

b) Now, let's apply this reasoning to genomic libraries. The maize haploid genome is 2×10^9 bp long. You have made a BAC library with an insert size of 2×10^5 bp. The library contains 10,000 clones. Would you expect that, somewhere in your library, every gene in the maize genome is represented? Explain your answer.

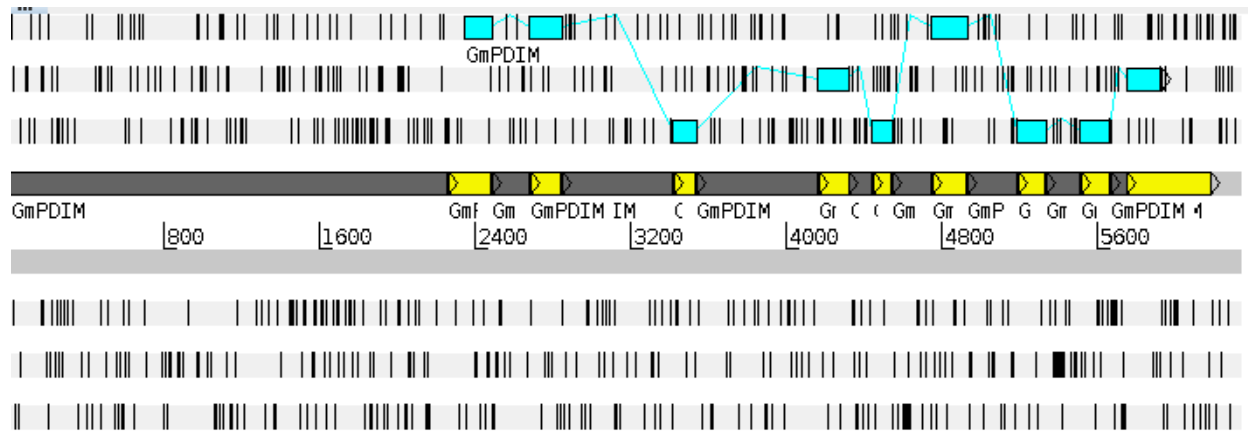
6. (10 points) Which do you think would yield more DNA: 100 mg of embryos, or 100 mg of tissue from fully expanded leaves?

Hint: Think about cell division and cell elongation as part of development.

7. (5 points) Media for co-cultivation of Agrobacterium with plant tissue needs Acetosyringone in order to induce Agrobacterium genes required for transformation. All other components are heat stable, but Acetosyringone would be destroyed by heat. How could you sterilize the acetosyringone so that it could be added to sterile media?

8. (10 points) The T_d for a 100 bp DNA fragment is 72°C at standard hybridization conditions. What would be the T_d for a 500 bp fragment, given similar conditions? Show your work.

9. (15 points) The image below depicts a clone for a gene for disulfide isomerase. In each of the 3 reading frames on the forward strand (top) and 3 reading frames on the reverse strand (bottom), stop codons are shown as vertical tick marks. A map of the gene is shown between the two strands.

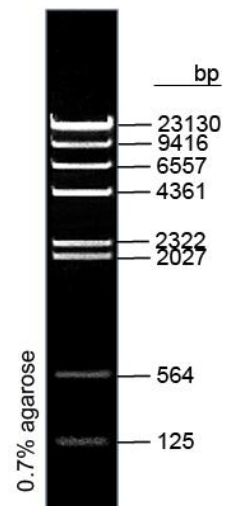


a) Which boxes are exons, and which are introns? What lets you distinguish between exons and introns in this diagram?

b) Is this from a cDNA library, or a genomic library?

c) The gene shown was cloned into the multiple cloning region of the pUC18 cloning vector. Many white colonies were screened, but the disulfide isomerase protein could not be detected in any of the clones. What is the most likely reason that the protein was not produced in any of the clones?

10. (5 points) The figure at right shows Lambda phage DNA digested with HindIII, separated by electrophoresis and stained with RedSafe. The length of each restriction fragment is indicated at right. Explain why some bands are brighter than others on the gel.



11. (10 points) Match statements a - e with one of the choices.

Enzyme	Cutting site
BamHI	G [^] GATCC
ApaI	GGGCC [^] C
NotI	GC [^] GGCCGC
AoxI	[^] GGCC
HaeII	RGCGC [^] Y

- a) Digestion with this enzyme gives the largest fragments
- b) Digestion with this enzyme gives the smallest fragments
- c) Enzymes whose overhangs are compatible for ligation
- d) Generates a 3' protruding end
- e) Has an asymmetric recognition sequence

Choices:

- 1. none
- 2. HaeII
- 3. NotI
- 4. AoxI
- 5. ApaI
- 6. BamHI
- 7. ApaI, AoxI
- 8. AoxI, NotI
- 9. ApaI, NotI

Genetic code

		Second base							
		U		C		A		G	
First base	U	UUU	Phe	UCU	Ser	UAU	Tyr	UGU	Cys
		UUC	Phe	UCC	Ser	UAC	Tyr	UGC	Cys
		UUA	Leu	UCA	Ser	UAA	STOP	UGA	STOP
		UUG	Leu	UCG	Ser	UAG	STOP	UGG	Trp
C	CUU	Leu	CCU	Pro	CAU	His	CGU	Arg	
	CUC	Leu	CCC	Pro	CAC	His	CGC	Arg	
	CUA	Leu	CCA	Pro	CAA	Gln	CGA	Arg	
	CUG	Leu	CCG	Pro	CAG	Gln	CGG	Arg	
A	AUU	Ile	ACU	Thr	AAU	Asn	AGU	Ser	
	AUC	Ile	ACC	Thr	AAC	Asn	AGC	Ser	
	AUA	Ile	ACA	Thr	AAA	Lys	AGA	Arg	
	AUG	Met	ACG	Thr	AAG	Lys	AGG	Arg	
G	GUU	Val	GCU	Ala	GAU	Asp	GGU	Gly	
	GUC	Val	GCC	Ala	GAC	Asp	GGC	Gly	
	GUA	Val	GCA	Ala	GAA	Glu	GGA	Gly	
	GUG	Val	GCG	Ala	GAG	Glu	GGG	Gly	

tRNA

UAC

5' AUG codon

mRNA

Frequencies of Restriction Sites (or other oligonucleotides)

length n	frequency: occurs every 4 ⁿ	example	sequence
1	4	Single nucleotide	G
2	16	Di-nucleotide	GT
3	64	Codon	ATG
4	256	Taq I	TCGA
5	1024	MbolI	GAAGA
6	4096	Hind III	AAGCTT
7	16384	Abe I	CCTCAGC
8	65536	Not I	GCGGCCGC