PLNT4610 BIOINFORMATICS

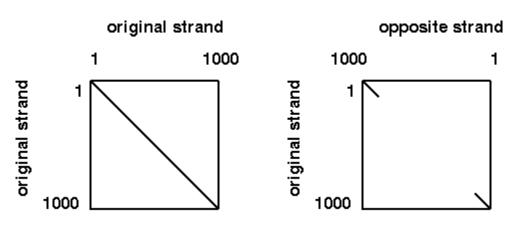
MID-TERM EXAMINATION

3:00 p.m. to 4:00 p.m. Monday, October 26, 2009

Answer any combination of questions totalling to <u>exactly</u> 100 points. If you answer questions totalling more than 100 points, answers will be discarded at random until the total points equal 100. This exam is worth 20% of the course grade.

Hand in this question sheets along with your exam book. All questions must be answered in the exam book. The exam sheets will be shreded after the exam.

- 1. (20 points) Define the following:
 - a) similar
 - b) homologous
 - c) analogous
 - d) orthologous
 - e) paralogous
- 2. (10 points) A sequence was compared with its opposite strand, showing short diagonals at each end. Explain this observation.



- 3. (20 points) There are many advantages to searching a protein sequence against a protein database, rather than searching a DNA sequence against a DNA database.
 - a) List two ways in which the protein search is faster
 - b) List two ways in which the protein search is more sensitive

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4. (5 points) It might seem trivial to generate the opposite strand of a sequence, so simple, in fact that you might be able to do it by a simple search and replace:

original sequence	AATCGTTTGCCCCCCTA
replace A with 1	11TCGTTTGCCCCCCT1
replace G with 2	11TC2TTT2CCCCCCT1
replace T with A	11AC2AAA2CCCCCA1
replace C with G	11AG2AAA2GGGGGGA1
replace 1 with T	TTAG2AAA2GGGGGAT
replace 2 with C	TTAGCAAACGGGGGGAT

What is the problem with this approach?

5. (15 points) A number of alternative genetic codes have been discovered. Examples are found in mitochondria, plastids, bacteria and archea. In all of the alternative genetic codes seen so far, most of the codons code for the same amino acids as in the Standard Genetic Code, with a few codons differing. For example in some cases, a stop codon codes for an amino acid, or a codon for an amino acid is used as a stop codon. In other cases, one or two codons are reassigned to a different amino acid.

Type of search	NCBI	FASTA
a)DNA vs. DNA database	blastn	fasta3 ssearch3 (slow, full Smith-Waterman alignment)
b) protein vs. protein database	blastp	fasta3 ssearch3 (slow, full Smith-Waterman alignment)
c) protein vs. translated DNA database	tblastn	tfasta3
d) translated DNA vs. translated DNA database	tblastx	tfastx3, tfasty3
e) translated DNA vs. protein database	blastx	fastx3, fasty3 (especially well-suited for cDNAs, which often contain frameshift errors)

Yeast mitochondria use a non-starndard genetic code. Suppose you had the sequences for a yeast mitochondrial gene, and its corresponding protein, and wished to find homolgues in other species.

How would the difference in genetic codes affect each of the types of searches listed above?

6. (10 points) The CLUSTAL/TCOFFEE family of programs begin creating a multiple alignment by building a guide tree based on the distances between sequences. Next, sequences are added to the alignment one at a time, and the alignment is adjusted to include each new sequence. What is the main shortcoming of this strategy?

7. (15 points)

For a given restriction endonuclease, how many possible restriction fragments can be generated

- a) in a complete digest of a linear molecule with n sites
- b) in a complete digest of a circular molecule with n sites
- c) in a partial* digest of a circular molecule with n sites
- *A partial digest means that if there are n sites, not all sites necessarily cut, resulting in a population of fragments with 1 cut, 2 cuts ... n cuts.
- 8. (5 points) In pairwise DNA sequence alignments, matches are scored as +1, mismatches as -1, and gaps as -2. Why are gaps more strongly weighted than mismatches?
- 9. (5 points) Describe what is meant by the term "E-value", for BLAST and FASTA database searches.
- 10. (10 points) Part of a multiple sequence alignment for PR10 defense genes from several species of pea is shown below. There are 5 paralogues of PR10, numbered 1 through 5, found in different pea species *Pisum sativum* (Ps), *Pisum humile* (Ph), *Pisum elatius* (Pe) and *Pisum fulvum* (Pf). (Note: blanks in the alignment indicate that the clone was truncated at the 5' or 3' end. In the case of Ypr10.Ps.2 blanks are also seen within the intron because that sequence comes from a cDNA clone. Thus, no intron sequence is available.). Which regions of the alignment would be best for designing gene-specific PCR primers:
 - •that would <u>only</u> work for a particular copy of the gene?
 - •that would be most likely to amplify <u>any</u> PR10 genes from more distant species from which PR10 had never been cloned before?

Notes on PCR primers: DNA synthesizers can create degenerate primers. For example, if some genes have A at a position, and other genes have G at that position, the primer pool will have two sets of primers with either A or G at that position. Thus, in the oligo specified as AYCCTCGTA, Y stands for purine. So two primers would be produced, AGCCTCGTA, and AACCTCGTA. In practical application, degeneracies can only be at a small number of positions in a primer.

	100	110	120	130	140	150	160	170	180 <i< th=""><th>190</th></i<>	190
1 Ypr10.Ps.1 1 Ypr10.Ph.1	ACTCCAAAGGTTAT	TGATGCCATCA	AAAAGTATCGA				CCATCAAGAAA A			
Ypr10.Pe.1 Ypr10.Pf.1	ACTCCAAAGGTTAT	TGATGCCATCA	\	AAT G						c.
2 Ypr10.Ph.2 Ypr10.Pf.2	ACTCCAAAGGTTAT ACTCCAAAGGTTAT	TGATGCCATCA	AAAGTATCGA	AAT.G			AA			c.
Ypr10.Ph.3 Ypr10.Pe.2 Ypr10.Pe.3	ACTCCAAAGGTTAT	IONIOCCHICA	mmomilo	.G		cc.c	. A			t .t
Ypr10.Pf.3 4 Ypr10.Ps.4	GTCCCAAAGGTGAT GTCCCAAAAGTTGT	CAAGGAAGCA(TGATTCAATCA	CAAGGAGTCGA AAGACTGTTGA	.G AAAT.A.C		CC.CG				t
	200	210	220	230 intron	240	250	260	270	280	290
F Ypr10.Ps.1	a-aatatnc-t-	-t-tt-acg	a-atat-c-t	-t-anta-ta	a-tannatt-t	tt-aa-t-t	tgnaatt-	ttntgt	-gcagATGGT	GAAAC
1 Ypr10.Ph.1 Ypr10.Pe.1	tta.a.g tta.a.g	gtt.	g	c.c.a	aa aa		C	a a		
Ypr10.Ps.2 2 Ypr10.Ph.2	tta.a.g	gtt.	g	c.c.a	aa		c	a		
Ypr10.Ps.3 Ypr10.Ph.3	.gt .gt	g	.ta. ta.	tc	.gttg.	t.tg.t.g.	agaa.c	aa.tg.g aa.gg.g	tC t	
Ypr10.Pe.3 Ypr10.Pf.3	.gt .g.gt	a	ta.	tc	.atta.	t.ta.c.a.	agaa.c	aa.gg.g	t t	
4 Ypr10.Ps.4 5 Ypr10.Ps.5									GAA	.A .C.G
	300 CAAGNATGTGTTGC	310 ACAAAGTGGAG	320 TTAGTAGATG	330 NTGCTAACTT	340 rggcttacaac	350 CTATAGCATAG	360 GTTGGNGGTGT	370 IGGANTTCCA	380 GACACAGTTG	390 AGA AG
1 Vpv10 Db 1	AC			m			m.	T C		
Ypr10.Pf.1	. AC			T			T	T T		
r ipriu.Ps.3	T A .			A	jAA		A			
3 Ypr10.Pn.3 Ypr10.Pe.2 Ypr10.Pe.3	TATATA.			A	GAAA GAAA		A	CG		
4 Ypr10.Ps.4	CTC.A.	C.A	2GCT	AAA	.T.G	CT	AAACC	AGC.A.AT	A.GTT.A.	A
	400 ATCTCATTNGAGGC	410	420 -CAG GACCAAA	430	440	450	460 AATATTACACA	470 AAAGGTGAT-	480 GCTGCTCC	490
1 Ypr10.Ph.1							C.T			G
Ypr10.Pe.1	T				A		C.T			C
2 Ypr10.Ph.2 Ypr10.Pf.2										C
Ypr10.Ph.3 Ypr10.Pe.2	GC	T					TC		AT	G G
Ypr10.Pf.3 4 Ypr10.Ps.4	G.TGCA.	A.TTA.T.TG.	A.GG	C.T	C.TT	A.ATC	TC		AT	G ATC
5 Ypr10.Ps.5										T
r Ypr10.Ps.1	500 AAGAGNAACTCAAG.	.CA		.GTCT			560 FNTGGCNNATC .TTC			
Ypr10.Pe.1 Ypr10.Pf.1	C	.CA								
2 Ypr10.Ph.2 Ypr10.Pf.2	C	.AA		.GTCT	GTC.T	.GGGA.TG.	.TTC	GAA.	C.A.A C.A.A	
Ypr10.Ps.3 Ypr10.Ph.3 Ypr10.Pe.2	GA GA	.AG	CATA	.GTAT	.GTC.T	.AGGA.TG.	.GTA	GAA.	C.A.A	
Ypr10.Pe.3 Ypr10.Pf.3 4 Ypr10.Ps.4	GA GA .T.CAGTT.GTG.T	GAAACAG	.CAAAC.	.GACA	GCA.A.	.AGGA.GT	.TAA	GGT.	A.T.GT	
	AGG.TG									